

HUMAN HEXOKINASE II : SEQUENCE AND HOMOLOGY TO OTHER HEXOKINASES*

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SUMMARY The amino acid sequence of human hexokinase II was deduced from the sequence of cDNA clones isolated from a skeletal muscle library. An open reading frame of 2751 bases encodes a protein of 917 amino acids. The deduced amino acid sequence has 94% identity with rat hexokinase II but only 72% identity with human hexokinase type I. In addition to hexokinase II clones, the human skeletal muscle cDNA library contained at least an equal number of clones of hexokinase I, the isoform reported to be typically found in kidney and brain. Genetic variation in hexokinase II could underlie insulin resistance in peripheral tissues and cause non-insulin-dependent diabetes mellitus. The availability of this sequence would facilitate investigating the role of mutations in the HKII gene in the etiology of this disease. © 1993 Academic Press, Inc.

The four mammalian hexokinases (ATP:D-hexose 6-phosphotransferase, EC 2.7.1.1.) catalyze the conversion of glucose to glucose-6-phosphate, the first step of intracellular glucose metabolism (1). Hexokinases I-III have a molecular weight of approximately 100,000, have a high affinity for glucose and are subject to a allosteric inhibition by glucose-6-phosphate (Glc-6-P). In contrast, hexokinase IV, also known as glucokinase, has a molecular weight of 50,000, a lower affinity for glucose, and is not feedback inhibited by physiological concentrations of Glc-6-P (2). The extensive sequence homology between the N- and C-terminal halves of hexokinases I-III suggests that the mammalian hexokinases evolved from an ancestral enzyme similar to glucokinase or the yeast hexokinase by gene duplication and tandem fusion (3 - 5). Type I hexokinase has been cloned from rat brain (6), bovine heart muscle (7) and from human kidney (8) and mouse hepatoma cells (9). Rat hexokinase II has been cloned from skeletal muscle and adipose tissue libraries (5, 10).

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Abbreviations: Glc-6-P, glucose 6-phosphate, NIDDM, non-insulin-dependent diabetes mellitus.

Hexokinase II is involved in the increased glucose uptake and utilization by adipose and skeletal muscle in response to insulin (11-13). Transcription of the hexokinase II gene in rat adipose and skeletal muscle is induced by insulin (5, 14). Non-insulin-dependent diabetes mellitus (NIDDM) is characterized by decreased glucose uptake and utilization by these tissues, and by relatively low basal levels of Glc-6-P (15, 16). Therefore, defects in HKII synthesis or activity may be implicated in the etiology of NIDDM. As a first step towards testing this hypothesis, we have isolated and sequenced HKII cDNA clones from a human skeletal library and deduced the complete amino acid sequence of the enzyme.

MATERIALS AND METHODS

A human (15 year old female Caucasian) skeletal muscle cDNA library (in bacteriophage λ gt 11) was purchased from Clontech Laboratories, Inc. (Catalog number HL 1124b, Palo Alto, CA). The full length rat hexokinase II cDNA probe (10) was kindly provided by Dr. John E. Wilson, Michigan State University. The human skeletal muscle cDNA library was screened for hexokinase II clones according to the protocol provided by the manufacturer. The probe was labeled by random priming using [32 P]dCTP to a specific activity of approximately 10^9 cpm/ μ g. The inserts in hybridizing clones were amplified by the polymerase chain reaction (17) using the following primers that are complementary to sequences flanking the Eco RI cloning site of λ gt 11:

Forward 5' GGTGGCGACGACTCCTGGAGCC 3'
Reverse 5' GACACCAGACCAACTGGTAATG 3'.

Thirty five cycles of 30 seconds of denaturation at 92°C, followed by 1 minute of annealing at 64°C, and 1-3 min. of extension at 72°C were employed using the Perkin-Elmer Cetus GeneAmp kit. Phage lysates (boiled) were used as templates for amplification reactions. The amplified inserts were directly sequenced as described (18) using the above two primers plus internal hexokinase II primers. Multiple alignment of hexokinase nucleotide and amino acid sequences were made using the GENALIGN program of IntelliGenetics, Inc. (Mountain View, CA).

RESULTS

Isolation of hexokinase cDNA clones. Forty two of the 3.5×10^5 phages in the human skeletal muscle cDNA library hybridized to the rat hexokinase II cDNA probe. The inserts in twenty of these clones were characterized by amplification and direct sequencing. The sequences of ten of these clones were identical to those found in human hexokinase I cDNA isolated from a kidney cDNA library (8). Seven of the 20 clones were found to have a high degree of nucleotide sequence homology with the rat hexokinase II cDNA sequence (10). These seven clones were used to determine the sequence of the coding region and parts of the untranslated regions of human hexokinase II cDNA. Based on the size of inserts in these clones, the human hexokinase II mRNA is predicted to be at least 4.8 kb in length.

Sequence of the human hexokinase II and homology to other hexokinases. The sequence of the entire coding region (both strands) and 117 and 273 nucleotides of the 5' and 3' untranslated regions, respectively is given in Figure 1. The sequence revealed an open reading frame of 2751 bases which encodes a protein of 917 amino acids. The calculated molecular weight of 102,365 closely resembles that deduced for rat hexokinase II (102,550)(5, 10) and human hexokinase I (102,519) (8). At the nucleotide level, the human and rat hexokinase II cDNA sequences are 87.2% identical, whereas human hexokinases I and II share 72.5% sequence identity. In the alignment shown in Fig. 2, the human and rat hexokinase II isozymes exhibit 94% sequence identity and 97% similarity with rat hexokinase II but only 72% identity and 87 % similarity with human hexokinase I.

As was observed for the rat hexokinases I, II, and III and human hexokinase I isozymes (8, 6,10, 20), there is striking homology between the N-terminal and C-terminal halves in human hexokinases I and II which are both homologous to the yeast hexokinase. Alignment of the two halves of hexokinases I and II indicates, respectively, 52% and 54% identity with a fusion point at amino acid residue 463, and that the C-terminal half lacks the first 15 N-terminal residues. It has been suggested that these N-terminal segments are important in anchoring these enzymes to the mitochondrial membrane (8, 21).

DISCUSSION

A significant finding in this study was that, in contrast to the extremely high ratio of hexokinase II to hexokinase I mRNA in rat skeletal muscle (5), clones encoding these two isoforms were observed to be equally represented in human skeletal muscle. This may have an important physiological implication since the synthesis of hexokinase II only is responsive to insulin action.

The striking sequence similarity between the N-terminal and C-terminal halves of hexokinase types I-III support the hypothesis advanced by several investigators that these isozymes evolved from hexokinase IV and yeast hexokinase-like enzymes (which have approximately half the molecular weight) by a process involving gene duplication followed by tandem ligation. The N-terminal half of rat brain hexokinase I contains the allosteric regulatory site for Glc-6-P (22), whereas the C-terminal half contains the catalytic site(23, 24). However, a recombinant C-terminal half of hexokinase I, expressed in *E. coli*, was shown to be both catalytically active and sensitive to inhibition by Glc-6-P (25). Putative amino acid residues involved in binding of glucose and of ATP have been deduced from the X-ray structure of yeast hexokinase and from comparison of sequences among the hexokinases (9, 20, 26-29). Residues corresponding to Ser-158, Asn-210, Glu-269, and Glu-302 (glucose binding domain) and Gly-X-Gly-X-X-Ala at 459-464

Figure 1. Nucleotide sequence of the cDNA and deduced amino acid sequence of human skeletal muscle hexokinase II.

HUM-HEXII	1	MIASH11AyfFeLNhdQVKVQDqLYhMRLSDETLLEIsRFRKEMEKGLGATHTPTAAVKMLPTFVRSTPDGTEHGEFLALDLGGTNFRV
RAT-HEXII	1	MIASHmIAcIFTELNgqVQVKVQDfLYhMRLSDETLLEIsRFRKEMEKGLGATHTPTAAVKMLPTFVRSTPDGTEHGEFLALDLGGTNFRV
HUM-HEXI	1	MIaql1AyyfTELKddQVKiDkYLYhMRLSDETLIdimRFRKEMKGLdfrnPTAcVKMLPTFVRSiPDGsEkGdFiALDLGGsFRi
consensus		MIASH11Ay-FeLN-dQVKVQDqLYhMRLSDETLLEIs-RFRKEMEKGLgattHTPTAAVKMLPTFVRSTPDGTEHGEFLALDLGGTNFRV
HUM-HEXII	93	LwVKTdNGLQKQVEMENQIYAIPEdIMRSGGTQLFDHIAECLANFMDKLIQIKdKKLP1LGFTFSFPCQTKLDESFLVSWTKGPKSSGVEGRD
RAT-HEXII	93	LRVrVTdNGLQrVEMENQIYAIPEdIMRSGGTQLFDHIAECLANFMDKLIQIKdKKLP1LGFTFSFPCQTKLDESFLVSWTKGPKSSGVEGRD
HUM-HEXI	93	LRVqVnhekQnVhMEseVYdtPeNtVhGSGGQLFDHIAECLdFMeKrkIKdKKLPvGPTFSFPCQqKiDeAiLitWTKrPKasGVEGdAd
consensus		LrV-VtdnglQ-VeMEngqIYAIPEdIMrSGGTQLFDHIAECLANFMDKLIQIKdKKLP1LGFTFSFPCQTKLDESfLvsWTKgPKsGVEGRD
HUM-HEXII	185	VVALIRKAIQRGGDFDIDIVAVVNDTVGTMMTCGYDDHNCIEGLIVGTGSNACYMEEMRHIDMVEGDEGRMCINMEWGAFGDDGsLNDIRTE
RAT-HEXII	185	VVDLIRKAIQRGGDFDIDIVAVVNDTVGTMMTCGYDDQNCIEGLIVGTGSNACYMEEMRHIDMVEGDEGRMCINMEWGAFGDDGtLNDIRTE
HUM-HEXI	185	VVKLnKAiKkGdyDanIVAVVNDTVGTMMTCGYDDqHCEVGLIIGTGTNACYMEELRHIDIVEGDEGRMCINtEWGAFGDDGsLeDIRTE
consensus		VV-LIRKAIqrGDFDIdIVAVVNDTVGTMMTCGYDDqNCIEGLIVGTGSNACYMEEMRHIDMVEGDEGRMCINMEWGAFGDDGsLnDIRTE
HUM-HEXII	277	FDqEIDMGSLNPGKQLFEKMSIGMYGELVRLILVKMAKeELLFgGKLSPELLnTGrFETKDiSDIEgeKDGIRKarevLMRLGLdPtQEDC
RAT-HEXII	277	FDREIDMGSLNPGKQLFEKMSIGMYGELVRLILVKMAKeELLFgGKLSPELLTGSFETKQVSDIEedKDGIEKAYqILMRGLnPlQEDC
HUM-HEXI	277	FDREIDrGSLNPGKQLFEKMSIGMYGELVRLILVKMAKegLLFeGritPELLTrGKFnTsDVSIAEKneKelnAkeILrLrGvePaddDC
consensus		FDREIDMGSLNPGKQLFEKMSIGMYGELVRLILVKMAKeELLF-GKLSPELLTtG-FETKdVSdIE--Kdgi-ka-eiLrMRGL-P-qeDC
HUM-HEXII	369	VATHRICQIVSTRASLCAATLAaVLqRIKENKGEERARSTIGVDSVYKkHPhfAKRLHKtVRRLPVpCDVRFLRSEDGSGKGAAMVTAVA
RAT-HEXII	369	VATHRICQIVSTRASLCAATLAaVLwRIKENKGEERLRSTIGVDSVYKkHPhfAKRLHkaVRRLPVpCDVRFLRSEDGSGKGAAMVTAVA
HUM-HEXI	369	VsvqhvCtIVSTRAnLVAATLgAilLnRlrdNKGTpRLRtTvGVDSYlYKtHPqysrRfHKtLRRLVPdsDVRFLISEGSGKGAAMVTAVA
consensus		VathricQIVSTRASLCAATLAaVL-RiKENKGEERLRSTIGVDSVYKkHPhfakRLHKtVRRLPdcDVRFLRSEDGSGKGAAMVTAVA
HUM-HEXII	461	YRLADQHRARQKTLelhLqLSHdQLLEVRRMKVEMERGLSKETHaSaAPVKMLPTTYVCATPDGTEKGDFLALDLGGTNFRVLLVRVRNGKwg
RAT-HEXII	461	YRLADQHRARQKTLesLkLSHeQLLEVRRMKVEMEGGLSKETHuAvAPVKMLPTTYVCATPDGTEKGDFLALDLGGTNFRVLLVRVRNGKtRG
HUM-HEXI	461	YRLAeQHRqieeTLahfhtKdmLLEVKKRMraEMEIGLrKgThnnAvVKMLPsFVrTPDGTENGDFLALDLGGTNFRVLLVkiRSgKKRt
consensus		YRLADQHRarqKTLelh-LshdQLLEVRRMKVEME-GLSKETHa-APVKMLPTyVoaTPDGTExGDFLALDLGGTNFRVLLVrVRNGK-rG
HUM-HEXII	553	VEMHNKIYAIpQEVmHGtGELFDHiaQCIADfLEYMGKGVSLPLGFTFSFPCQqNSLDsILLKWTGFKFASGCCGEdVATLKEAIHRR
RAT-HEXII	553	VEMHNKIYsIpQEVmHGtGELFDHIVQCIADfLEYMGKGVSLPLGFTFSFPCQqNSLDqSILLKWTGFKFASGCCGEdVVTLLKEAIHRR
HUM-HEXI	553	VEMHNKIYAIPIEImGtGELFDHIVeCISDFLDYMGIKGprrMLGFTFSFPCQqSLDagLlItWTKGFKAtcdVghDVVTLLrdaIKRR
consensus		VEMHNKIYAIpQEVmHGtGELFDHivQCIADfLEYMGKGVsLPLGFTFSFPCQqNSLD-sILLKWTGFKFASGCCGEdVvTLLKEAIHRR
HUM-HEXII	645	EEFDLDVVAVVNDTVRTMMTCGYEDPhCEVGLIVGTGSNACYMEEMRNVELVdGEEGRMCVNMWGAFGDNGCLDDIRTePDVADELleft
RAT-HEXII	645	EEFDLDVVAVVNDTVGTMMTCGYEDPhCEVGLIVGTGSNACYMEEMRNVELVdGEEGRMCVNMWGAFGDNGCLDDIRTVFDVADELsLNP
HUM-HEXI	645	EEFDLDVVAVVNDTVGTMMTCaYDePtCEVGLIVGTGSNACYMEEMRNVEhVdGdgGqMCINMEWGAFGDNGCLDDIRThYDrIvNysSLNa
consensus		EEFDLDVVAVVNDTVGTMMTCgyEdPhCEVGLIVGTGSNACYMEEMrNVELVdGeeGrMcVNMWGAFGDNGCLDD-RT-fdVaDELsLNP
HUM-HEXII	737	GKQRFEMKISGMYLGEIVRNILIDFTKRGLLFRGRISERLKRIGIFETKFLSQIESDCIALLQVRAILqHGLSTCCDdSIIVKEVCTVVAR
RAT-HEXII	737	GKQRFEMKISGMYLGEIVRNILIDFTKRGLLFRGRISERLKRIGIFETKFLSQIESDCIALLQVRAILrHGLSTCCDdSIIVKEVCTVVAR
HUM-HEXI	737	GKQRYEKMSIGMYLGEIVRNILIDFTKRGfLFRGgISetmKTRIGIFETKFLSQIESDrLALLQVRAILqgLGhLStCCDdSIIVKtCVVgsR
HUM-HEXII	829	RAAQLCGAGMAAVVDrIRENRGLDnLVTVGVdGTLYKLHPHFAKVMHETVrDLAPKCDVSFLSLEDGSGKGAALITAVACRIReAGQR
RAT-HEXII	829	RAAQLCGAGMAAVVDKIRENRGLDnLVTVGVdGTLYKLHPHFAKVMHETVrDLAPKCDVSFLSLEDGSGKGAALITAVACRIReAGQR
HUM-HEXI	829	RAAQLCGAGMAAVVDKIRENRGLDrLnVTVGVdGTLYKLHPHfScrImHqTVkeLSPKCnVSFLSLEDGSGKGAALITAVGvRIReass
consensus		RAAQLCGAGMAAVVDKIRENRGLD-LkTVGVdGTLYKLHPHFAKVMHETVrDLAPKCDVSFL-LEDGSGKGAALITAVACRIReagqr

Figure 2. Comparison of the deduced amino acid sequences of human HKI, HKII, and rat HKII. Multiple alignment of human hexokinase II (HUM-HEXII), rat hexokinase II (RAT-HEXII), and human hexokinase I (HUM-HEXI). The vertical lines denote matches between pairs. The consensus sequence shows exact matches in uppercase letters and the consensus character in lowercase letters. Horizontal lines indicate no consensus.

(ATP binding domain) of the yeast hexokinase are all conserved in both halves of human and rat hexokinases I and II (residues 448-453 and 896-901). Botstein and colleagues (30) identified the following six missense mutations in the yeast

hexokinase II gene that significantly affected the function of this enzyme: Gly89Asp, Thr190Ile, Gly235Cys, Asn237His, Ser306Phe, and the double mutant Gln367Glu and Ile381Met. The residues corresponding to the first five of these positions are fully conserved in both halves of hexokinases I and II. The Gln residue at position 376 is conserved only in the corresponding position 365 of the N-terminal domain of hexokinase II, and the Ile residue is conserved only in the corresponding position 370 of the C-terminal domain of hexokinase II, suggesting that neither residue is important in catalysis.

Missense mutations in the human type IV hexokinase (glucokinase) gene have been identified in patients with an autosomal dominant form of NIDDM (31). All of these mutations (at residues 70, 175, 182, 203, 228, 256, 261, 279, 300, 299, 309, and 414) cause changes in catalytic properties. The residues in both halves of hexokinases I and II found at sites that correspond to all but two (279 and 175) of these mutated sites in hexokinase IV are conserved. Arg and Met are substituted for Glu at the position corresponding to residue 279 (hexokinase II 283) in only the C-terminal domains of hexokinases I and II, respectively. The residue 279 mutant allele encodes an enzyme with a higher K_m for glucose (but not ATP) and lower V_{max} (30). Thus, the apparent absence of catalytic activity in the N-terminal domain may be at least partially due to substitution at residue 279. Residue 179 in hexokinase I and hexokinase II corresponding to Gly 175 in hexokinase IV is conserved in both halves except for the presence of Asp in the C-terminal domain of hexokinase I. The Gly175Arg mutation in hexokinase IV causes a reduction in V_{max} and an increase in K_m for glucose (30). It appears, therefore, that the enzyme can tolerate substitution of an acidic but not a basic residue at this position.

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